Study on Genetic Variability, Association and Classification of F₄ and F₅ Progenies of Groundnut (*Arachis hypogaea* L.) for Late Leaf Spot Reaction, Yield and its Component Traits

N. SHILPA¹, N. MARAPPA², C. R. JAHIR BASHA³, D. L. SAVITHARMMA⁴ AND R. L. RAVIKUMAR⁵ ^{1,2&4}Department of Genetics and Plant Breeding, ³Department of Plant Pathology, ⁵Department of Plant Biotechnology, College of Agriculture, UAS, GKVK, Bengaluru - 560 065

e-Mail : shilpanmurthy100@gmail.com

AUTHORS CONTRIBUTION

N. SHILPA :

Conceptualization, design, manuscript writing and analysis of data; N. MARAPPA & C. R. JAHIR BASHA : Conceptualization, design, editing and supervision; D. L. SAVITHARMMA & R. L. RAVIKUMAR : Conceptualization, design, material preparation and supervision

Corresponding Author :

N. SHILPA Department of Genetics and Plant Breeding, College of Agriculture, UAS, GKVK, Bengaluru

Received : August 2023 Accepted : October 2023

Abstract

In the present study, F₄ and F₅ populations were evaluated to study the genetic variability for traits related to per cent disease incidence (PDI) for late leaf spot (LLS) reaction, pod yield and its component characters. Higher estimates of PCV and GCV were recorded for PDI at all the intervals, number of pods per plant, kernel yield per plant and pod yield per plant. High heritability accompanied with high GAM was observed for pods per plant, kernel yield per plant, pod yield per plant while plant height showed moderate GAM. Pods per plant, kernel yield per plant, sound mature kernel per cent and shelling per cent exhibited strong positive association with pod yield, whereas PDI@60 DAS, PDI@75 DAS, PDI@90 DAS and PDI@105 DAS exhibited significant negative association. Kernel yield exerted highest positive direct effect on pod yield. PDI at all the intervals recorded negative direct effect on pod yield. Indirect effect of number of pods per plant and kernel weight were high for pod yield. Principal component analysis (PCA) indicted that first and second component justified 58.4 per cent of variations among different traits. Biplot analysis of PCA results aided in identifying 15 LLS resistant lines. Kernel weight per plant has direct effect on pod weight per plant. Therefore, selecting this trait will increase pod yield. The identified lines can be evaluated in varied rainfed environments to exploit their disease resistance and yield potential and the validated lines can be utilised as improved cultivars.

Keywords : Association, Groundnut, Genetic variability, Late leaf spot disease, Principal component analysis

G_{ROUNDNUT} (*Arachis hypogaea* L.) is one of the principal oil seed crops of the world. It is popularly known as 'King of oil seeds' because of its high edible oil content and it is utilized for human consumption as vegetable oil and protein and as fodder for livestock. It contains edible oil (44-53%), protein (23-25%), carbohydrate (20%) and fibre (3%). It is also rich in calcium, phosphorus, iron, thiamine (B1), riboflavin (B2) and niacin. Groundnut, being majorly grown in rainy season, is affected by several pests and diseases leading to lower productivity. Among several diseases affecting groundnut, late leaf spot disease caused by *Phaeoisariopsis personata* results in reduction in pod and haulm yield of 25.3 and 53.0 per cent, respectively (Eswara Reddy and Venkateshwara Rao, 1999). Many methods have been suggested to control LLS including combination of cultural and chemical measures, but they were of limited success. Genetic studies on LLS resistance suggested that resistance to this fungal disease is complex and polygenic in nature and sensitive to environments (Sujay *et al.*, 2012). It has been long known that environment factors, like temperature and humidity are important affecting infection and development of LLS disease. Land races and wild species of groundnut possess high level of resistance to foliar diseases, but the resistance is generally linked to low productivity, late maturity, poor adaptability and undesirable pod features (Wynne et al., 1981 and Singh and Mass, 1982). Germplasm originating from secondary centers of diversity were resistant to foliar diseases with desirable agronomic backgrounds, but their productivity levels were low (Singh and Mass, 1982). Therefore, development of new varieties that are tolerant to LLS and high yielding cultivars suitable for cultivation is the need of the hour. In plant breeding, development of any variety requires suitable breeding method and for success of any crop improvement programme requires genetic variability in the initial breeding population is required. For practicing selection in the population, presence of genetic variability is a pre-requisite. For developing appropriate breeding strategy, information on phenotypic and genotypic variation for pod yield and its component traits is needed in order to practice selection. Breeders often use segregating generations as source population and exercise selection for identifying homozygous lines with better performance with a view to develop varieties. But pod yield being complex trait is dependent on a number of other traits. The traits that influence yield and their direct and indirect effects can be studied through correlation and path analysis. This becomes important in order to select the traits that indirectly influence yield. Therefore, a study was undertaken to estimate the genetic variability, heritability, correlation and path analysis for LLS reaction and pod yield related traits.

MATERIAL AND METHODS

The material used in the experiment consisted of two segregating populations of F_4 and F_5 generations, developed from the cross TMV 2 × GPBD 4. TMV 2, female parent of the cross is highly susceptible to LLS, whereas GPBD 4 is resistant to LLS. The initial objective of the study was to screen the population for LLS resistance and identify resistant/tolerant sources for LLS. Ninety four lines of F_4 generation were screened in *kharif* 2021 for LLS reaction and its yield component traits. Later the lines were forwarded to F_5 generation through progeny test and observations on pod yield related traits were recorded in summer

2022. The present investigation was conducted at experimental plots (K-block), Department of Genetics and Plant Breeding, College of Agriculture, GKVK, University of Agricultural Sciences, Bangalore. In *kharif* season F_4 population was grown in separate blocks along with checks and parents in augmented design as suggested by Federer (1959) and observations were recorded on each of the F₄ individuals. TMV 2 was used as a spreader row, since the variety is highly susceptible to LLS disease to create natural epiphytotic condition for the spread of the disease. It was replicated after an interval of six lines. LLS reaction was scored following modified 9 point scale for LLS disease given by Subrahmanyam et al. (1995) on 60th, 75th, 90th and 105th days after sowing till the crop reached physiological maturity. The data was analysed in R software ver. 4.2.1. Genotypic and phenotypic coefficients of variability were computed both in F_4 and F_5 generation for each character as per the method suggested by Burton and Devane (1953). Heritability was estimated in F_4 and F_5 generation for each character using the formula given by Lush (1945). Genetic advance was estimated using the formula of Johnson et al. (1955). Correlation coefficients were computed amongst the pairs of characters using the formula given by Panse and Sukhatme (1964). Principal Component analysis and biplot analysis were computed and biplot graph was plotted using XLSTAT © 2023. The lines were categorised based on the response into four groups (A, B, C and D) for LLS resistant and susceptibility.

RESULTS AND DISCUSSION

Analysis of Variance (ANOVA) for LLS reaction, pod yield and its component traits of F_4 and F_5 generations of the cross TMV 2 × GPBD 4 is presented in Table 1 and Table 2. Mean sum of squares genotypes were significant for all the traits under study in F_4 population except for traits such as PDI@90 DAS, plant height and kernel yield per plant. In F_5 population, all traits studied were significant, except kernel yield. Analysis of variance for both progenies and checks displayed significantly greater difference for characters like PDI@60 DAS, PDI@75 DAS, PDI@105 DAS, days to first flowering, number of

	S	
	ŭ	
	20	
	2	
ζ	ろ	
	g	
	Þ	
	t_{l}	
	-	
	2	
	Ľ	
	1	
	h,	h
	-	U.
-	4	۲
-	Ř	
,	MA_{5}	
-	01A9	
-	u ot A	
1 1 1	ial of As	
	nal of As	
	urnal of As	
1 1 1	wrnal of As	
	ournal of As	
1 1 1 1	Journal of As	
1 V 1	e Journal of As	
1 U I	re Journal of As	
	ore Journal of As	
I 1 1 1	sore Journal of A	
1 U I I U I	vsore Journal of As	
1 U I	Avsore Journal of As	
1 V I I V I	Mvsore Journal of As	

							TABLI	[]]						
Mean sum	of squ	ares of l	F ₄ progen	ies deri	ved fron	n the cro	ss TMV	$2 \times \text{GPB}$	D 4 for pc	od yield a	nd its attril	buting tra	its in grour	ndnut
Source	Df	PDI@ 60 DAS	PDI@ 75 DAS	PDI@ 90 DAS	PDI@ 105 DAS	DFF	Hd	PBP	NPP	PWP	NKP	KWP	SMK	SP
Treatment (ignoring Blocks)	96	0.55 *	1.5 **	2.39	2.88 **	4.68 **	10.93	34.96 **	83.68 **	30.69 *	320.6 *	** 12.01	68.76 **	173.08 **
Treatment: Check	7	4.61 **	22.01 **	23.47	23.94 **	22.23 **	9.75	2	360.29 **	255.55 **	1152.51 *	** 115.04 *	* 111.5 **	148.24
Treatment: Test vs. Check	1	15.96 **	6.12 **	1.57	0.06	0.04	0.43	14.61 **	21.24	200.54 **	121.75	97.61 *	* 3.84	0.09
Treatment: Test	93	0.3	1.01 *	1.94	2.46 **	4.35 **	11.07	35.89 **	78.4 **	24.03 *	304.85 *	** 8.87	68.54 **	175.48 **
Block (eliminating Treatments)	9	0.27	0.26	0.53	0.28	0.54	9.51	4.92 *	53.13	22.5	198.51 *	20.1 *	22.87	131.3 *
Residual	12	0.2	0.36	0.35	0.15	1.07	5.18	1.13	18.19	9.22	48.81	5.97	12.47	43.32
Df = Deg PWP = 1 Mean sum	od wei, od wei, of squ	reedom, I ght per pla ares of F	DFF = Days ant (g), NK SP SP SP SP SP SP SP SP SP SP SP SP SP	P = Num = Shellin ies deri	lowering, ber of kerr ng percent ved from	PH = Plan nel per pla age (%), * 1 the cro	tt height (c mt, KWP = Significan TABLF ss TMV	m), PBP = = Kernel w(t @P = 0.0 i 2 × GPB]	Primary bra eight per pla 5, **Signifi D 4 for po	anches per unt (g), SM cant @P = cant @P a	olant, NPP = K = Sound m 0.01 0.01 od its attrik	Number o ature kerne buting tra	f pods per pla el <i>per cent</i> (% its in grour	nt,), idnut
Source			Df	DFF	KW	VP	NKP	Ê	P	BP	Hd	PWP	SMI	X SP
Treatment (ignorin	g Bloc	ks)	96	19.05 *	× ×	56	373.35 **	* 99	.4 ** 3	.07 * 1	6.77 **	28 *	120.12 **	214.33 **
Treatment: Check			7	3.83	109.0	04 ** 1	396.99 **	* 458.8	81 **	2.1	5.09 28	85.18 **	177.81 **	123.7
Treatment: Test vs.	. Check		1	8.81	103.4	44 **	20.33	15.3	34 0	.03 10	8.93 **	96.69 **	524.75 **	0.4
Treatment: Test			93	19.49 *	5.	38	355.13 **	* 92.5	57 ** 3	.12 * 1	6.03 **	21.73	114.53 **	218.58 **
Block (eliminating	Treatn	nents)	9	2.02	13.′	* 62	150.74	38.5	57 5	.32 *	6.09	14.04	26.63	141.45 *

Df = Degrees of freedom, DFF = Days to first flowering, PH = Plant height (cm), PBP = Primary branches per plant, NPP = Number of pods per plant, PWP = Pod weight per plant (g), NKP = Number of kernel per plant, KWP = Kernel weight per plant (g), SMK = Sound mature kernel *per cent* (%), SP = Shelling percentage (%), *Significant @P = 0.05, **Significant

40.26

11.02

9.52

3.6

1.18

22.27

54.69

3.91

6.26

12

Residual

pods per plant, pod yield per plant, number of kernel per plant, kernel yield per plant, sound mature kernel per cent except plant height, primary branches per plant and shelling percentage in F_4 generation. Mean sum of squares of checks versus progenies also recorded significant difference for all the characters except PDI@90 DAS, PDI@105 DAS days to first flowering, plant height, number of pods per plant, sound mature kernel *per cent* and shelling *per cent*. Non-significant variation was observed for kernel yield per plant for mean sum of squares of progenies in F₅ generation and there was no significant variation in case of progenies versus checks for shelling per *cent* in both the generations. The results obtained are in accordance with Bhavya et al. (2017) who recorded non-significant variation for traits pods/ plant and shelling *per cent* in progenies of F_4 generation and non-significant variation for days to first flowering and shelling *per cent* in F₅ population of the cross GKVK-16 \times KCG-2. ANOVA revealed presence of significant variation for most of the characters under study among progenies except for kernel yield per plant thus indicating presence of sufficient amount of genetic variability for the traits under consideration.

Genetic Variability Parameters

Variability refers to presence of a wide range of phenotypic and genotypic differences among the genotypes of the population. These differences can be due to the differences in genetic constitution of the individuals or due to environment influence on the traits. In order to carry out selection in any population presence of variability is a must. Therefore, knowledge about the magnitude of genetic variability in a given population is of prime importance to plant breeders. Information of phenotypic and genotypic coefficients of variation provides the actual variance of the characters. Heritability and genetic advance estimation provides information about the genetic gain when selection is applied. In this study, variability parameters such as phenotypic (PCV) and genotypic coefficients of variation (GCV), heritability and genetic advance over per cent mean (GAM) were estimated.

The range of variation was wide for all the characters under study. Important pod yield component traits such as number for pods per plant recorded a wide range of 9.9 to 49.5 and 8.8 to 47.6 in F_4 and F_5 population, respectively. Traits such as pod yield per plant recorded range of 4.5 g to 28.6 g in F_4 generation and 3.6 g to 26.5 g in F_5 generation. Sound mature kernel *per cent* ranged between 44.6 to 89.8 per cent and 33.7 to 86.0 per cent in F_4 and F_5 population, respectively.

Estimates of genetic variability revealed high PCV and GCV for PD1@75 DAS (27.7 and 22.2 %), PD1@90 DAS (29.9 and 27.1 %) and PD1@105 DAS (28.1 and 27.3 %). Similar results of high PCV and GCV were recorded by John et al. (2006) for late leaf spot disease. Medium GCV estimates were recorded for PD1@60 DAS. Days to first flowering and plant height exhibited low to medium PCV and GCV estimates. Ganesan and Sudhakar (1995), John et al. (2006), Raut et al. (2010), Hiremath et al. (2011), Vishnuvardhan et al. (2012) and Zongo et al. (2017) also reported moderate estimates of PCV and GCV for plant height. Number of pods per plant, pod yield per plant, number of kernel per plant and shelling per cent recorded high estimates of PCV and GCV in F₄ and F_5 generations. The present findings were in accordance with results obtained by Venkataravana et al. (2000), Khote et al. (2009), Patil et al. (2014), Hyndavi (2015), Padmaja et al. (2015), Kadam et al. (2017), Patel (2017), Zongo et al. (2017) and Jambagi and Savithramma (2020) for pod yield and its component traits. Sound mature kernel percentage recorded medium to high PCV and GCV estimates in both the generations. Hugar and Savitramma (2015) recorded high PCV and GCV estimates for sound mature kernel percentage. Presence of high phenotypic and genotypic variation indicates that there is sufficient amount of variability present in the population and selection can be practiced. Lower estimates of PCV and GCV indicates lower magnitude of variation for these traits. Selection will be ineffective when the trait is not heritable or less heritable. Therefore, information on the magnitude of trait heritability is of prime importance for groundnut crop improvement. In this study, high broad sense heritability and genetic advance over per cent mean (GAM) was recorded for PD1@75 DAS (64.3 and 36.7 %), PD1@90 DAS (82.2 and 50.8 %), PD1@105 DAS (93.8 and 54.5%), primary branches per plant, number of pods per plant, number of kernel

per plant, sound mature kernel *per cent* and shelling *per cent* in both F_4 and F_5 generations. A similar trend was noticed by John *et al.* (2006) and Giri *et al.* (2009). Verma *et al.* (2002), John *et al.* (2007), Raut *et al.* (2010), Hyndavi (2015), Kumar *et al.* (2016) and Zongo *et al.* (2017) who reported high heritability coupled with high GAM for number of branches per plant, number of pods per plant, pod yield per plant and kernel yield. Medium heritability and GAM was recorded for PD1@60 DAS (32.0 and 13.7 %), plant height (53.2 and 10.5 %) and kernel yield per plant (32.7 and 26.4 %) in F_4 generation, whereas medium and low heritability was exhibited by pod yield per plant (56.2 and 37.1 %) and kernel yield per plant

(27.3 and 16.8 %) in F_5 generation, respectively. The present results were in accordance with results obtained by Jayabose *et al.* (2022) who observed moderate genetic advance for kernel yield. Medium GAM was recorded by days to first flowering (19.5 %), plant height (18.1 %) and kernel yield per plant (16.8 %) in F_5 generation (Table 3).

Correlation Analysis

Correlation is the association between the traits and the information about correction allows breeders to practice selection for suitable traits that improve yield. Traits that show positive correlation can be selected in the breeding program for improvement of yield.

TABLE 3

Estimates of genetic variability parameters for late leaf spot disease reaction, pod yield and its component traits in F_4 and F_5 generations of cross TMV 2 × GPBD 4

Trait	Generation	Mean ± SE	Maximum	Minimum	GCV (%)	PCV (%)	h2bs (%)	GA	GAM (%)
PDI@ 60 DAS	F ₄	$2.64\pm\ 0.06$	3.7	1.9	11.7	20.8	32.0	0.4	13.7
PDI@ 75 DAS	F_4	$3.63 \pm \ 0.11$	5.6	2.3	22.2	27.7	64.3	1.3	36.7
PDI@ 90 DAS	F_4	$4.66\pm\ 0.14$	7.6	3.5	27.1	29.9	82.2	2.4	50.8
PDI@ 105 DAS	F_4	5.57 ± 0.16	8.3	4.1	27.3	28.1	93.8	3.0	54.5
DFF	F_4	$32.43 \pm \ 0.22$	36.5	27.9	5.6	6.4	75.4	3.2	10.0
	F ₅	$31.7\pm\ 0.16$	35.9	28.6	11.5	14.0	67.9	6.2	19.5
PH	F_4	$34.86\pm\ 0.36$	42.3	26.8	7.0	9.5	53.2	3.7	10.5
	F ₅	35.3 ± 1.56	45.6	27.7	10.0	11.3	77.5	6.4	18.1
PBP	F_4	$7.76 \pm \ 0.61$	10.9	4.6	76.0	77.2	96.9	12.0	54.3
	F ₅	6.8 ± 0.36	10.6	3.4	20.6	26.2	62.1	2.3	33.5
NPP	F_4	$7.34\pm\ 0.88$	49.5	9.9	28.4	32.4	76.8	14.0	51.3
	F ₅	$28.7~\pm~1.23$	47.6	8.8	29.3	33.6	76.0	15.1	52.6
PWP	F_4	13.99 ± 0.49	28.6	4.5	27.5	35.0	61.6	6.2	44.6
	F ₅	14.6 ± 0.78	26.5	3.6	24.0	32.0	56.2	5.4	37.1
NKP	F_4	51.74 ± 1.74	94.6	16.9	30.9	33.7	84.0	30.3	58.5
	F ₅	$54.5~\pm~0.27$	89.9	15.6	31.8	34.6	84.6	32.9	60.4
KWP	F_4	8.7 ± 0.39	15.8	2.1	22.4	39.1	32.7	2.0	26.4
	F ₅	$12.6~\pm~1.67$	15.6	2.4	15.6	29.8	27.3	1.3	16.8
SMK	F_4	75.83 ± 0.81	89.9	44.7	9.9	10.9	81.8	14.0	18.4
	F ₅	$71.6~\pm~0.25$	86.1	33.7	14.2	15.0	90.4	20.0	27.9
SP	F_4	55.3 ± 1.64	82.1	30.4	20.8	24.0	75.3	20.6	37.2
	F ₅	55.5 ± 1.23	87.2	33.6	24.1	26.6	81.6	24.9	44.8

DFF = Days to first flowering PH = Plant height (cm) PBP = Primary branches per plant NPP = Number of pods per plant

PWP = Pod weight per plant (g) NKP = Number of kernel per plant KWP = Kernel weight per plant (g)

SMK = Sound mature kernel *per cent* (%) SP = Shelling percentage (%)

N. SHILPA *et al*.

Thus correlation studies help in indirect selection of characters that affect yield.

PD1@60 DAS, PD1@75 DAS, PD1@90 DAS and PD1@105 DAS recorded significant negative association with traits such as number of pods per plant, pod weight per plant, number of kernel per plant, kernel yield per plant, sound mature kernel per cent and shelling per cent, whereas PD1@75 DAS exhibited non-significant negative correlation for sound mature kernel per cent in both the generations. Days to first flowering, plant height and primary branches per plant recorded negative and nonsignificant association with PDI at all the intervals. Similar results were reported by Vasanthi et al. (1998) who recorded negative association of late leaf spot with days to first flowering. Present results are also in accordance with Giri et al. (2009) who observed negative significant correlation between late leaf spot severity (%) and pod yield related traits in groundnut genotypes. Similarly, Yashaswini et al. (2021) recorded significant negative correlation of PDI at all the intervals to pod yield related traits in F_4 generation of groundnut cross TMV 2 × ICGV 86699. Important pod yield component traits such as number of plants, number of kernel per plant, kernel weight per plant and sound mature kernel per cent recorded significant positive correlation for pod yield per plant in both F₄ and F_5 generations (Table 4). The present results were in accordance with Lakshmidevamma et al. (2004), Dandu et al. (2012), Nandini and Savitramma (2012), Prabhu et al. (2014) and Jayabose et al. (2022) where they observed positive significant association between kernel yield, number of pods per plant, pod yield and shelling percentage. Shelling per cent exhibited nonsignificant positive correlation for number of pods per plant (0.1818), pod yield per plant (0.0991) and number of kernel per plant (0.1717) in F_4 generation. Similar results were reported by Shoba et al. (2012) and Prabhu et al. (2015).

Shelling *per cent* recorded negative non-significant correlation for number of pods per plant (0.1074) and number of kernel per plant (-0.1295), whereas negative significant association was seen between shelling *per cent* and pod weight per plant (-0.2713) in F_5

generation. Days to first flowering, plant height and primary branches per plant had non-significant positive correlation with number of pods per plant, pod weight per plant and kernel weight per plant. In the study conducted by Jayabose *et al.* (2022), pod yield and days to flowering, number of pods and kernel weight, shelling percentage and pod yield were negatively correlated.

Path Analysis

Correlation analysis provides the mutual relationship between various plant characters and determines the component character on which selection can be based for genetic improvement in yield. Path analysis splits the correlation coefficients into direct and indirect effects. Path analysis showing direct and indirect effects can be employed to get high simultaneous selection response for several characters from a diverse population. Therefore, path coefficient analysis could provide a more realistic picture of the interrelationship (Memon *et al.*, 2019).

Direct Effects on Pod Yield Per plant

Number of pods per plant, kernel weight per plant, sound mature kernel per cent and shelling per cent recorded positive direct effect on pod yield per plant in both F_4 and F_5 population (Table 5). The results indicate that selection for these traits will positively improve pod yield. Similar results were reported by Lakshmidevamma et al. (2004) and Giri et al. (2009). In F₄ generation, PDI@60 DAS (-0.0007), PDI@75 DAS (-0.0271), PDI@90 DAS (-0.5542) and PDI@105 DAS (-0.2600) exhibited negative direct effects on pod yield per plant. Selection of genotypes with low PDI will be suitable in order to increase pod yield. In the study conducted by Giri et al. (2009), late leaf spot disease severity exerted negative direct on pod yield per plant. Other studies conducted by Sumanthi and Muralitharan (2007) and Shoba et al. (2012), Kushwah et al. (2016), Jahanzaib et al. (2020) and Javabose et al. (2022) supported the above results.

Indirect Effects on Pod Yield Per plant

Indirect effect of number of pods per plant on pod yield per plant is significantly positive in both F_4

The Mysore Journal of Agricultural Sciences

Association analysis for LLS disease reaction. prowth parameters, nod vield and its attributing traits in TABLE 4

PDI@ 60 DAS F ₁ 1 0.7016 *** 0.5707 *** 0.5354 -0.6356 0.083 -0.7136 *** -0.5319	/P NKP	KWP SMK	SP
	*** -0.4738 *** -0.	.5454 *** -0.2325 *	-0.2929 *
	*** -0.5979 *** -0.	.5300 *** -0.2923	-0.1354 *
	*** -0.5032 *** -0.	.4080 *** -0.2412 *	* -0.0218 *
	*** -0.5860 *** -0.	.4965 *** -0.3627 *	* -0.1190 *
F_1 I 0.0158 0.0544 0.0844 0.0434 0.0132 PH F_1 I 0.0745 0.1482 0.1188 0.1248 0.0354 FB F_1 0.0745 0.1482 0.1188 0.1248 0.1350 PBP F_1 0.0745 0.1180 0.0148 0.1350 0.01616 0.0248 0.0248 0.0248 0.0248 0.0128 0.0128 0.0128 0.01261 0.0248	-0.0085 0.		0.2031 *
PH F_4 0.1248 0.1258 0.1024 0.1268	-0.0484 -0.0	.0753 -0.0497	0.0083
F3 I 0.1350 0.0584 0.1170 0.0616 -0.268 P3P F_4 0 0.0556 -0.048 0.0473 -0.1024 F3 F3 0 0.0556 -0.048 0.0473 -0.1024 P3P F_4 0 0.0556 -0.048 0.0473 -0.1024 N3P F_4 0.1750 1 0.0556 -0.048 0.0763 -0.1750 N4P F_4 0.1760 1 0.0556 -0.1495 0.0143 -0.1750 N4P F_4 0.1760 0.0564 0.0135 0.0924 -0.1750 0.7852 N4P F_4 0.166 0.0564 0.0132 0.0132 0.0132 -0.1750 N4P F_4 0.0141 0.0164 0.0132 -0.143 0.0123 0.0142 -0.0164 N4P F_4 0.0124 0.0124 0.0123 0.0126 0.0124 -0.0164 K4W F_4 F_4 F_4 F_4 F_4 -0.0164 -0.0164 -0.0164 -0	0.1248 0.	.1952 -0.0317	0.1265
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	0.0616 -0.	.2268 * 0.1245	-0.4439 **:
F_3 1 -0.1566 -0.1455 -0.1688 -0.1750 NPP F_4 1 0.9153 0.9442 *** 0.7855 F< 1 0.9153 0.9442 *** 0.7855 PWP F_4 1 0.9153 0.9442 *** 0.7956 PWP F_4 1 0.9153 *** 0.7944 *** 0.7944 NKP F_4 1 0.8649 *** 0.7944 *** 0.7944 NKP F_4 1 0.8649 *** 1 0.9182 *** 0.7844 KWP F_4 1 0.8649 *** 1 0.7863 *** 0.7844 KWP F_4 1 0.8649 *** 1 0.7863 *** 0.6631 KWP F_4	0.0473 -0.	.1024 0.005	-0.1207
NPP F_4 0.99153 0.9942 *** 0.7036 F_5 F_5 0.9024 *** 0.7036 PWP F_4 0.9024 *** 0.7036 PWP F_4 0.9024 *** 0.7036 PWP F_6 0.9024 *** 0.7044 NKP F_6 0.9024 *** 0.7044 NKP F_6 0.7044 0.7044 0.7044 NKP F_6 0.7044 0.7044 0.7304 SMK F_6 0.7044 0.7044 0.7804 SMK F_6 0.7804 0.7944 0.7804 F_6 F_6 0.7804 0.7804 0.7804 0.7804 F_7 F_8 F_9 0.6640 F_8 0.6640 F_8 0.6640 F_8 0.7804 F_8	-0.1688 -0.	.1750 -0.1678	-0.1372
$ \begin{array}{llllllllllllllllllllllllllllllllllll$	*** 0.9942 *** 0.	.7855 *** 0.6164 *	** 0.1818
	*** 0.9924 *** 0.	.7096 *** 0.8634 *	** -0.1074
F5 0.8687 *** 0.6632 NKP F4 0.7804 1 0.7804 KWP F5 1 0.7804 1 0.6810 KWP F4 1 0.7804 1 0.6810 KWP F4 1 1 0.6810 1 SMK F4 1 </td <td>0.9182 *** 0.</td> <td>.7944 *** 0.5447 *</td> <td>** 0.099</td>	0.9182 *** 0.	.7944 *** 0.5447 *	** 0.099
NKP F 1 0.7804 F E 0.6810 0.6810 KWP F 0.6810 0.6810 SMK F 0.6810 0.6810 SMK F 0.6810 0.6810 SMK F 0.6810 0.6810	0.8687 *** 0.	.6632 *** 0.7700 *	** -0.2713 **
F5 0.6810 KWP F4 SMK F4 SMK F4	1 0.	.7804 *** 0.6336 *	** 0.1717
KWP E ₄ F ₅ SMK F ₄ F ₅	0.	.6810 *** 0.8837 *	** -0.1295
SMK F ₅ F ₅		1 0.3554 *	** 0.6500 ***
SMK F ₄ F ₅		1 0.5000 *	** 0.4625 ***
т. г.		1	0.0441 **
5			0.2928 **
SF F4			1
ъ s			1

Mysore J. Agric. Sci., 57 (4) : 298-310 (2023)

N. SHILPA *et al*.

i nenetypie put		traits in	F_4 and F_5	generation	ns of cross	TMV 2×0	GPBD 4	y lota alla te	suttrouting
Traits	Gen.	NPP	KWP	SMK	PDI@60 DAS	PDI@75 DAS	PDI@90 DAS	PDI@105 DAS	PWP
NPP	F_4	0.0016	1.0011	0.0002	0.0004	0.0023	-0.0008	-0.0005	0.5056 **
	F ₅	1.3033	0.2008	0.0152	-	-	-	-	0.4951 **
KWP	F_4	1.0042	0.0013	0.0013	0.0022	0.0091	-0.0008	0.0016	0.4043 *
	F_5	0.0013	0.0086	0.0060	-	-	-	-	0.1094 *
SMK	F_4	0.0047	-0.0002	0.0111	0.0031	-0.0001	0.0012	0.0058	0.0838 **
	F ₅	0.0013	0.0071	0.0281	-	-	-	-	0.2616 *
PDI@60 DAS	F_4	0.0026	0.0014	-0.0001	-0.0007	0.0030	-0.0002	0.1747	-0.1952 **
PDI@75 DAS	F_4	-0.2839	0.0005	0.0223	0.1324	-0.0271	0.0001	0.0841	-0.0943 *
PDI@90 DAS	F_4	-0.0736	-0.2452	0.0085	-0.0123	0.0030	-0.5542	-0.0084	-0.9315 *
PDI@105 DAS	F_4	-0.1359	-0.0856	0.0378	0.0649	0.0555	0.0820	-0.2600	-0.5341 *

TABLE 5 Phenotypic path coefficient analysis indicating direct and indirect effect on LLS, pod yield and its attributing

DFF = Days to first flowering, PH = Plant height (cm), PBP = Primary branches per plant, NPP = Number of pods per plant, PWP = Pod weight per plant (g), SP = Shelling percentage (%), NKP = Number of kernel per plant, *Significant @P = 0.05, KWP = Kernel weight per plant (g), **Significant @P = 0.01, SMK = Sound mature kernel *per cent* (%)

(0.5056) and F₅ (0.4951) generations. Number of pods per plant recorded positive indirect effect on pod weight per plant through kernel weight per plant (1.0011 and 1.3013) and sound mature kernel per cent (0.0002 and 0.0152) in F₄ and F₅ generations. Kernel weight per plant exhibited positive indirect effect through number of pods per plant (1.0042 and 1.3116) and sound mature kernel per cent (0.0013 and 0.0152) in F_4 and F_5 populations. In the study conducted by Jayabose et al. (2022), number of pods had indirect effect on number of pods per plant contributing to kernel yield. Similarly, Patil et al. (2014) and Pachauri and Sikarwar (2022) recorded moderate indirect effect on kernel yield through number of pods. In conclusion, the traits kernel yield per plant, number of pods per plant exhibited positive direct effect on pod yield per plant. LLS disease incidence recorded significant negative indirect effect on pod yield per plant. Giri et al. (2009) in their study recorded negative indirect effect on pod yield per plant. Therefore, selection should be based on number of pods per plant and kernel yield per plant is more effective. Selection for low PDI will aid in development of lines resistance LLS and to increase pod yield in the population.

Principal Component and Biplot Analysis

Principal Component analysis is a method to find the linear combination that accounts for as much as variability as possible. In the present study principal component (PC) analysis was carried out to group genotypes of groundnut into different categories based on the mean performance for reaction to LLS. For the classification of lines on the basis of PDI@90, biplot was drawn using XLSTAT *ver*. 2023. Genotypes were grouped into different groups and is displayed in biplot of PCA1 and PCA2 (Fig. 1)

Combination of PC1 and PC2 explained highest amount of variation of 58.4 per cent (Table 6), mainly distinguishing the lines into four groups, out of which group C had high PDI@90 DAS. Other groups showed strong correlation between kernel weight per plant and shelling percentage. Significant correlation recorded for number of pods per plant, pod weight per plant and sound mature kernel percentage. Genotypes were categorised into four groups (A, B, C and D) based on their disease reaction.

 $Group A-resistant \ lines \ with \ low \ 90 @PDI \ DAS \ with \ high \ shelling \ percentage \ and \ kernel \ weight \ per \ plant.$



Biplot (axes PC1 and PC22: 58.54 %)

Fig. 1 : Grouping of the genotypes and identification of LLS resistant lines using Biplot analysis

TABLE 6
Estimates of eigen values and percentage of
variation in F_4 population of cross TMV2 × GPBD4
in groundnut

		0	
Principal component values	Eigen value	Variability (%)	Cumulative %
PC1	4.579	45.795	45.795
PC2	1.275	12.750	58.544
PC3	1.065	10.647	69.192
PC4	0.950	9.495	78.687
PC5	0.923	9.226	87.913
PC6	0.759	7.589	95.501
PC7	0.310	3.105	98.606
PC8	0.119	1.194	99.800
PC9	0.017	0.174	99.974
PC10	0.003	0.026	100.000

The Mysore Journal of Agricultural Sciences

Group B – moderately resistant and had high pod number, pod weight and SMK *per cent*. Group C – susceptible lines with high 90@PDI and group D with moderate susceptibility and is graphically represented in biplot. Our results showed that lines *viz.*, 13, 16, 19, 28, 32, 36, 43, 45, 47, 54, 61, 67, 68, 78 and 83 were highly resistant and high yielding (Table 7 and Fig. 2).

Similar result was reported in safflower by Bahrami and Karimi (2014) in Tomato by Thi (2016) and Suresh *et al.* (2018) and in Groundnut by Savita *et al.* (2014) and Jambagi *et al.*, 2020. In our study, high GCV, h² and GAM was observed for pod yield and its component traits which indicate additive gene action and is amenable for selection. Since characters *viz.*, number of pods per plant, kernel weight per plant and SMK *per cent* had significant positive effect on pod weight per plant, these traits can be considered as

Genotype	DFF	PH	PBP	NPP	PWP	NKP	KWP	SMK	SP
13	32.4	28.6	8.5	37.6	21.5	73.5	13.5	82.6	62.79
16	34.3	34.6	6.9	42.9	22.7	81.3	14.5	83.1	63.88
19	29.8	41.3	10.4	45.7	21.4	89.6	15.6	83.9	72.90
28	34.5	39.6	8.5	40.6	19.5	76.9	13.2	81.0	67.69
32	31.2	35.6	4.8	32.7	15.5	61.2	7.5	79.2	48.39
36	31.2	34.5	7.5	42.1	20.3	80.4	12.6	78.0	62.07
43	34.6	32.4	7.5	26.5	14.5	50.6	8.5	76.1	58.62
45	31.5	36.5	8.5	20.4	11.5	38.5	6.68	68.8	58.09
47	32.4	36.5	7.2	28.5	16.5	55.5	8.9	74.8	53.94
54	30.4	41.2	9.3	38.6	21.5	74.5	13.2	82.1	61.49
61	30.6	36.7	6.4	37.8	23.7	73.2	15.8	82.7	66.67
67	34.5	38.3	6.1	41.4	23.6	79.5	12.6	82.4	53.39
68	31.2	30.5	5.9	38.7	22.5	74.6	10.5	81.0	46.67
78	34.2	38.7	6.8	42.3	23.7	81.5	10.9	82.0	45.99
83	32.8	32.7	6.3	49.5	28.6	94.6	14.5	84.1	50.78

TABLE 7 Promising genotypes from the cross TMV2 \times GPBD4 for LLS resistance and pod yield related traits

DFF = Days to first flowering PH = Plant height (cm) PBP = Primary branches per plant NPP = Number of pods per plant PWP = Pod weight per plant (g) NKP = Number of kernel per plant KWP = Kernel weight per plant (g) SMK = Sound mature kernel *per cent* (%) SP = Shelling percentage (%)



Fig. 2 : Classification of the genotypes into different disease reaction

important selection criteria for improving pod yield in groundnut. The resistant genotypes identified could be used as a potential source of resistance to LLS in groundnut improvement programme.

REFERENCES

- BAHRAMI, F., ARZNI AND KARIMI, V., 2014, Evaluation of yield based drought tolerance indices for seeing safflower genotypes. *Agron. J.*, **106** (4) : 1219 - 1224.
- BHAVYA, M. R., SHANTHALA, J., SAVITHRAMMA, D. L. AND SYED SAB, 2017, Variability, heritability and association studies in F_4 and F_5 generation for traits related to water use efficiency and yield traits in groundnut (*Arachis hypogaea* L.). *Plant Archives*, **17** (2) : 1353 - 1360.
- BURTON, G. K. AND DEVANE, E. M., 1953, Estimating heritability in tall Fescue (*Festuca arundinaceae*) from replicated clonal material. *Agron. J.*, **45** : 478 481.
- DANDU, S. J., VASANTHI, R. P., REDDY, K. R. AND SUDHAKAR, P., 2012, Inheritance studies of plant height, pod and seed attributes in F_2 generation of certain ground nut (*Arachis hypogaea* L.) crosses. *Int. J. Appl. Biol. Pharm. Tech.*, **3** (1): 419 - 424.
- ESWARA REDDY, N. P. AND VENKATESWARA RAO, K., 1999, Chemical control of *Phaeoisariopsis* leaf spot of groundnut. J. Plant dis. Prot., **106**: 507 - 511.

- FEDERER, W. T., 1959, Augmented designs. *Hawaiian Planters Record*, **55** : 191 - 208.
- GANESAN, K. AND SUDHAKAR, D., 1995, Variability studies in Spanish bunch groundnut. *Madras Agric. J.*, **82**: 395 - 397.
- GIRI, R. R., TOPROPE, V. N. AND JAGTAP, P. K., 2009, Genetic variability, character association and path analysis for yield, its component traits and late leaf spot, *Phaeoisariopsis personata* (Berk and curt), in groundnut. *Int. J. Plant Sci.*, 4 (2): 551 - 555.
- HIREMATH, C. P., NADAF, H. L. AND KEERTHI, C. M., 2011, Induced genetic variability and correlation studies for yield and its component traits in groundnut (*Arachis hypogaea* L.). *Elect. J. Plant Breed.*, 2 (1) : 135 - 142.
- HUGAR, A. AND SAVITHRAMMA, D. L., 2015, Genetic variability studies for yield and surrogate traits related to water use efficiency in the recombinant inbred line (RIL) population derived from NRCG 12568 x NRCG 12326 of groundnut (Arachis hypogaea L.). Int. J. Agric. Sci. Res., 5: 321 - 328.
- HYNDAVI, Y., 2015, Genetic variability studies in F_4 and F_5 populations of selected crosses for traits related to water use efficiency, pod yield and its components in groundnut (*Arachis hypogaea* L.). *M.Sc. Thesis*, Univ. Agric. Sci., Bangalore (India).
- JAHANZAIB, M., NAWAZ, N., RSHAD, M., KHURSHID, H., HUSSAIN, M. AND KHAN, S. A., 2020, Genetic variability, traits association and path analysis in advanced lines of groundnut. *J. Innov. Sci.*, 7 (1): 88 - 97.
- JAMBAGI, B. K. P. AND SAVITHRAMMA, D. L., 2020, Genetic variability for physiological traits and pod yield in groundnut RILs under different water stress conditions. *Mysore J. Agric. Sci.*, **54** (4) : 90 - 96.
- JAMBAGI, B. K. P., SAVITHRAMMA, D. L., CHAUHAN, S. AND KUNDA, S., 2020, Identification of drought tolerant recombinant inbred lines in groundnut based on drought tolerant indices. J. Pharmacogn. Phytochem., 6 (14): 145 - 167.
- JAYABOSE, KANNAPPAN, P. L., VISWANATHAN, N., MANIVANNAN AND RAJENDRAN, L., 2022, Variability, correlation and path analyse in segregating population of groundnut (*Arachis hypogaea* L.). *Elect. J. Plant Breed.*, **13** (3) : 1099 - 1100.

- JOHN, K., MURALI KRISHNA, T., VASANTHI, R. P. RAMAIAH, M., 2006, Variability studies in groundnut germplasm. *Legume Res.*, **29** (3) : 219 - 220.
- JOHN, K., VASANTHI, R. P. AND VENKATESWARLU, O., 2007, Variability and correlation studies for pod yield and its attributes in F_2 generation of six Virginia x Spanish crosses of groundnut (*Arachis hypogaea* L.). *Legume Res.*, **30** (4) : 292 - 296.
- JOHNSON, H. W., ROBINSON, H. F. AND COMSTOCK, R. E., 1955, Estimates of genetic and environmental variability in soybeans. *Agron. J.*, **47** (7) : 314 - 318.
- KADAM, P. S., DESAI, D. T., JAGDISH, U., CHAUHAN, D. A.
 AND SHELKE, B. L., 2017, Variability, heritability and genetic advance in groundnut. J. Maharashtra Agric. Univ., 32 (1): 71 73.
- KHOTE, A. C., BENDALE, V. W., BHAVE, S. G. AND PATIL, P. P., 2009, Genetic variability, heritability and genetic advance in some exotic genotypes of groundnut (*Arachis hypogaea* L.). Crop Res., 37 (1, 2 & 3): 186 191.
- KUMAR, S. R., SEKHAR, M. R., DUTTA, S. S., SINGH, S. D. AND VERMA, S. K., 2016, Evaluation of 15 F_2 crosses for variability, heritability and genetic advance in groundnut (*Arachis hypogaea* L.). *Environ. Ecol.*, **34** (4C) : 2425 - 2430.
- KUSHWAH, A., GUPTA, S., SHARMA, S. R. AND PRADHAN, K., 2016, Genetic variability, correlation coefficient and path coefficient analysis for yield and component traits in groundnut. *Indian J. Ecol.*, **43** (2): 85 - 89.
- LAKSHMIDEVAMMA, T. N., GOWDA, M. AND MAHADEVU, P., 2004, Character association and path analysis in groundnut (*Arachis hypogaea* L.). *Mysore J. Agric. Sci.*, **38** (2) : 221 - 226.
- LUSH, J. L., 1945, Heritability of quantitative characters in farm animals. Proc. 8th Cong. *Genet. Herieditas.*, **35**: 356 - 375.
- MEMON, J. T., KACHHADIA, V. H., DEDANIYA, A. P. AND RUMIT, P., 2019, Character association and path analysis in F_2 generation of groundnut. *Int. J. Chem. Stud.*, 7 (3): 1329 - 1334.

- NANDINI, C. AND SAVITHRAMMA, D. L., 2012, Character association and path analysis in F_o recombinant inbred line population of the cross NRCG 12568 \times NRCG 12326 in groundnut (Arachis hypogea L.). Asian J. Bio. Sci., 7 (1): 55 - 58.
- PACHAURI, P. AND SIKARWAR, R. S., 2022, Correlation and path analysis of different environments for yield and component traits in groundnut (Arachis hypogaea L.). J. Pharm. Innov., 11 (2): 1181 - 1186.
- PADMAJA, D., ESWARI, K., RAO, B. M. AND PRASAD, S. G., 2015, Genetic variability studies in F, population of Groundnut (Arachis hypogeaea L.). Helix., 1 : 668 - 672.
- PANSE, V. G. AND SUKHATME, P. V., 1964, Statistical methods for agricultural research workers. ICAR, New Delhi, pp.: 381.
- PATEL, C. K., 2017, Genetic variation and interrelationship studies in F₂ generations of groundnut (Arachis hypogaea L.). M. Sc. Thesis, Junagadh Agric. Univ., Junagadh (India).
- PATIL, A. S., PUNEWAR, A. A., NANDANWAR, H. R. AND SHAH, K. P., 2014, Estimation of variability parameters for vield and its component traits in groundnut (Arachis hypogaea L.). Bioscan., 9:749 - 754.
- PRABHU, R., MANIVANNAN, N., MOTHILAL, A. AND IBRAHIM, S. M., 2014, Magnitude and direction of association for yield and yield attributes in groundnut (Arachis hypogaea L.). Elect. J. Plant Breed., 5 (4): 824 - 827.
- PRABHU, R., MANIVANNAN, N., MOTHILAL, A. AND IBRAHIM, S. M., 2015, Estimates of genetic variability parameters for yield and yield attributes in groundnut (Arachis hypogaea L.). Int. J. Agri. Environ. Biotech., 8 (3): 729 - 737.
- RAUT, R. D., DHADUK, L. K. AND VACHHANI, J. H., 2010, Studies on genetic variability and direct selections for important traits in segregating materials of groundnut (Arachis hypogaea L.). Int. J. Agric. Sci., **6**:234 - 237.
- SAVITA, S. K., KENCHNAGOUDAR, P. V. AND NADAF, H. L., 2014, Genetic variability for drought tolerance in advanced

breeding lines in groundnut. Karnataka J. Agric. Sci., 27 (2) : 116 - 120.

- SHOBA, D., MANIVANNAN, N. AND VINDHIYAVARMAN, P., 2012, Correlation and path coefficient analysis in ground nut (Arachis hypogaea L.). Madras Agric. J., 99 (1-3) : 18 - 20.
- SINGH, A. K. AND Moss, J. P., 1982, Utilisation of wild relatives in genetic improvement of Arachis hypogaea L. Theor. Appl. Genet., 61: 305 - 314.
- SUBRAHMANYAM, P., MCDONALD, D., WALIYAR, F., REDDY, L. J., NIGAM, S. N., GIBBONS, R. W., RAO, V. R., SINGH, A. K., PANDE, S. AND REDDY, P. M., 1995, Screening methods and sources of resistance to rust and late leaf spot of groundnut. Information Bulletin, No. 47; ICRISAT: Patancheru, India, pp. : 24.
- SUJAY, M. V. C., GOWDA, M. K., PANDEY, R. S., BHAT, Y. P., KHEDIKAR, H. L., NADAF, B., GAUTAMI, C., SARVAMANGALA, S., LINGARAJU, T., RADHAKRISHAN, S. J., KNAPP, R. K. AND VARSHNEY, 2012, Quantitative trait locus analysis and construction of consensus genetic map for foliar disease resistance based on two recombinant inbred line populations in cultivated groundnut (Arachis hypogaea L.). Mol. Breed., 30 : 773 - 788.
- SUMANTHI, P. AND MURALIDHARAN, V., 2007, Character association and path coefficient analysis in confectionery type groundnut (Arachis hypogaea L.). Madras Agric. J., 94 (1/6): 105 - 109.
- SURESH, K., 2018, Genetic analysis for fruit yield and yield components and tagging traits related to water use efficiency in tomato (Solanum spp.). Ph.D. Thesis, Univ. Agric. Sci., Bangalore.
- THI, N. N., 2016, Phenotypic diversity and association mapping for drought resistance and fruit yield in cultivated and related species of tomato (Solanum Spp.). Ph. D. Thesis, Univ. Agril. Sci., Bengaluru.
- VASANTHI, R. P., NAIDU, P. H. AND RAO, A. S., 1998, Genetic variability and correlation of yield component traits and foliar disease resistance in groundnut. J. Oilseeds Res., 15 (2): 345 - 347.

- VENKATARAVANA, P., SHERIFF, R. A., KULKARNI, R. S., SHANKARANARAYANA, V. AND FATHIMA, P. S., 2000, Correlation and path analysis in groundnut (*Arachis hypogaea* L.). *Mysore J. Agric. Sci.*, **34** (4): 321 - 325.
- VERMA, Y. P. A. K., HAIDER, Z. A. AND MAHTO, J. L., 2002, Variability studies in spanish bunch groundnut (Arachis hypogaea L.). J. Res. Birsa Agric. Univ., 14 (1):91-93.
- VISHNUVARDHAN, K. M., VASANTHI, R. P., REDDY, K. H. P. AND REDDY, B. V., 2012, Genetic variability studies for yield attributes and resistance to foliar diseases in groundnut (*Arachis hypogea* L.). *Int. J. Appl. Biol. Pharm. Technol.*, **3**: 390 - 394.
- WYNNE, J. C. AND GREGORY, W. C., 1981, Peanut breeding. Advances in Agronomy, **34** : 39 - 72.
- YASHASWINI, R., 2021, Assessment of genetic variability in F_3 and F_4 populations for late leaf spot disease resistance, pod yield and its attributing traits in selected crosses of groundnut. *M. Sc Thesis*, Univ. Agric. Sci., Bangalore.
- ZONGO, A., NANA, A. T., SAWADOGO, M., KONATE, A. K., SANKARA, P., NTARE, B. R. AND DESMAE, H., 2017, Variability and correlation among groundnut populations for early leaf spot, pod yield and agronomic traits. *Agron.*, **52** (7) : 1 - 11.